



Bursaphelenchus hofmanni Braasch, 1998 growth substrate in hops nurseries

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To date, seven *Bursaphelenchus* species have been reported in surveys of the Czech Republic (Čermák *et al.*, 2013). However, the occurrence of some species, such as *Bursaphelenchus hofmanni* Braasch, 1998, is limited to single detections in imported coniferous wood (unpubl. data in Braasch, 2001). During a survey to determine pathogenic agents on hops (*Humulus lupulus* L.) conducted by State Phytosanitary Administration (CZ) in hops fields and nurseries in 2012, *B. hofmanni* was found in a mixture of peat and soil in a hops seedling nursery in Žatec (Bohemia, Czech Republic). The occurrence of *Bursaphelenchus* species in non-woody plants or substrates has been previously reported for species of the *fungivorus* group, such as *B. gonzalezi* Loof, 1964, *B. hunti* Giblin & Kaya, 1983 and *B. fungivorus* Franklin & Hooper, 1962. Species belonging to the *hofmanni* group are often associated with wood products such as packaging material (Gu *et al.*, 2006) and, to our knowledge, this is the first time that this species has been found associated with peat substrate and soil.

Nematodes were isolated from 60 g of peat substrate (3 parts peat to 1 part soil) associated with hops seedlings

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Short communication

Bursaphelenchus associated with peat in the Czech Republic

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Using the Baermann funnel technique. *Bursaphelenchus* were killed and formalin and transferred to pure glycerin Grisse (1969). Two mature females and used to establish a culture maintained in *Helianthus scaber*, growing on 5% malt extract agar

Identification was confirmed by both morphological analyses (sequencing of the SSU and LSU rDNA loci). DNA was extracted as follows: specimens were collected into 20 µl of extraction buffer (10 mM Tris-HCl, pH 8.8; 1 mM EDTA; 0.5% Triton X-100 (v/v); 100 µg ml⁻¹ Proteinase K) in a microfuge tube. Each sample was ground using a pestle and mortar and incubated at 55°C for 1 h and subsequently at 100°C for 10 min. The mixture was used as a template for PCR. The SSU, LSU and ITS regions were amplified using a Phusion High Fidelity DNA polymerase (having 3' → 5' exonuclease activity). The DNA polymerase (Promega) was added to the reaction mixture and incubated at 72°C for 10 min for elongation step to create an A-tailed PCR product. The primer used

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